The Isolation and Identification of Active Compound of *Dendrophthoe praelonga* (Blume) Miq. Extract Against Breast Cancer Cells (MCF-7)

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**ABSTRACT**

Petai mistletoe (*Dendrophthoe praelonga* (Blume) Miq) is semi-parasitic plant. Parasite plant has been used traditionally in Indonesia to prevent or treat degenerative diseases, such as cancer. The purpose of this study was to indentify anticancer compound of petai mistletoe leaves extract and to examine its antioxidant activity (based on DPPH method) and cytotoxic activity on breast cancer (MCF-7) cell lines in vitro using Alamar blue method.

This research aims to isolating, characterize, determine antioxidant activity and cytotoxic activity. Separation was conducting column chromatography and characterize was conducted using TLC, FT-IR, LC-MS and NMR-1D. From the result obtained, the isolated compound is flavonol quercetine (3,3,7,3',4' pethahydroxyflavone) which active antioxidant with IC₅₀ value of 9.4 μg/mL and the value of cytotoxic activity with IC₅₀ 750.62 μg/mL.

**INTRODUCTION**

Cancer is a degenerative disease that potentially causes death (Torre et al. 2016). The increased amount of cancer occurrences is triggered by many factors, such as lifestyle, distribution patterns of population, and economic development (Jemal et al. 2011). Breast cancer is a kind of cancer that causes death in 14% of 30% of breast cancer sufferers among women (Siegel et al. 2017). One of the efforts in the development of thorough therapies is needed to overcome breast cancer and to reduce the mortality rate of the sufferers (Hanahan et al. 2011).

Chemotherapy techniques in the treatment of cancer have several disadvantages, such as the resistance to particular classes of the cytotoxic drug, side effects, and financial issues in the treatment process (Pearce et al. 2017). Accordingly, it indicates that this kind of cancer therapy has not been efficient, so it is needed for the researchers to develop a chemopreventive agent that can afford both effective and efficient values. Chemopreventive is an agent that potentially inhibits the development of cancer cells, and is able to reduce the growth of abnormal cells, and to reverse the process of carcinogenesis (Ko & Moon 2015).

A compound, that has the potential as a chemopreventive agent, is able to reduce the risk of cancer by impeding the stages in the initiating preneoplastic lesions by a source of carcinogens. To obtain compounds that have potential as chemopreventive agents are to explore natural materials, especially from some plants (Desai et al, 2008). One of the triggers of cancer can be caused by the free radicals compound that attacks cells of the human body. These free radical compounds are thought to be an initial factor in the onset of cancer including breast cancer (Risky & Suyatno, 2014). The antioxidant compound is a material that is able to constrain and prevent oxidation of an ingredient or compound that is easily oxidized by radical on an ongoing basis, and is able to reduce oxidative stress (Dai & Mumper 2010).
According to the belief in Indonesian people that the parasitic plant *Dendrophthoe pentandra* (L.) Miq is believed to have potential as an anticancer agent with a low toxicity value, and is likely to be an antioxidant (Artani et al. 2012). This is in line with the research conducted by Gamal and Septananda (2013) that parasites that grow on cottonwood plants (*D. petandra*) are able to suppress p53 mutants in HeLa cells in vitro. In addition, based on research conducted by Wicaksono & Permana (2013) stated that ethanol extracts from mango parasites (*D. petandra*) can cure the tissue structure in terms of colon cancer. In this study we reported isolation and identification of active compound from the leaves of the petai parasite (*Dendrophthoe praelonga* (Blume) Miq), as well as carrying out bioactivity testing such as antioxidant testing by using DPPH method, and cytotoxic testing of breast cancer cells (MCF-7) in vitro by using the alamar blue method. Active compound from extract mistletoe based on analysis of the compound using mass spectroscopy, Infra Red and NMR.

**METHODS**

**Plant material**
The plant material is mistletoe (*Dendrophthoe praelonga* (Blume) Miq) grow on petai (*Perikia speciosa*) tree in Cilegon, Banten, and determination was conducted at Herbarium Bogoriense Bogor

**Extraction**
Leaves *D. praelonga* powder (850 g) were macerated at room temperature with n-hexane for 24 hours (3x2L) and repeated then the residue was macerated with ethylacetate and ethanol 96%. The filtrates were evaporated under vacuum to obtain a gummy residue.

**Antioxidant Activity Test**
Antioxidant analysis was conducted using DPPH (1,1-diphenyl-2 picrylhydrazyl) free scavenging activity (Molyneux, 2004). Various concentration sample diluted in 8 mL methanol (p.a) mixed with 2 mL of methanol solution containing DPPH, result in a final concentration of DPPH of 0,1 mM and sample concentration up to 100 μg/mL. The mixture was shaken and left for 30 minutes at room temperature. The absorbance was then measured using spectrophotometer at 515 nm.

**Isolation and Purification of Ethanol Extract**
The fractionation was conducted using silica gel column chromatograph using increasing solvent polarity (n-hexane, ethyl acetat, and methanol). Activity guided isolation was conducted antioxidant activity using DPPH method (free radical scavenging activity). Fractiions with high antioxidant activity were purified by rinsing methanol a water. Thin Layer Chromatography was conducted throught out the fraction and purification process.

**Chromatography Profile of Sample**
Fractions from the ethanol extract were analyzed using thin layer chromatography (TLC). Each sample was eluted using plat GF254 and ODS as stationary phase n-hexane: ethyl acetat (7:3), methanol: ethyl acetat (8:2), and methanol: water (8:2) as mobile phase.

**Preparation of cancer cell line**
The cancer cell line used in this research was breast carcinoma (MCF-7). The cells was cultured in RPMI Medium with FBS 10%. The cells was cultured at temperature 37° with 95% water content and 5% CO2 for 3 days until the cells cultures become confluent 60-70%. After washing with new medium, it was an incubation again for 24h. The culturs were then washed with PBS 1-2 times and were suspended using typsin-EDTA solution. The cells that have been suspended added with new media.

**In vitro anticancer assay**
Toxicity testing of cancer cells using Alamar blue method. The cells line of 100 µL of breast cancer cells (MCF-7) added with 10 µL of test solution with a concentration variation of 62.5; 125; 250; 500; and 1000 µg / ml. The sample was incubated for 24 hours at 37°C. The coloring process is performed by adding a blue alamar solution for 4 hours. Color intensity was measured by ELISA (Thermo Fisher Scientific) at a wavelength of 560 & 590 nm.

**Statistical Analysis**
All analytical values shown represent the means of three repilcates, were analyzed using one-way ANOVA by SPSS 16.0 (Statistical Package for the Social Sciences) for Windows. Mean separation test between treatments was performed using Duncan’s multiple range test. *P* value ≤ 0.05 was considered statistically significant.
RESULTS AND DISCUSSION

Total yield extracts

The plant material (*D. praelonga*) were collected and dried. The plant extracts were collected with different solvents (*n*-hexane, ethyl acetate and ethanol). The final yield of leaves extracts in different solvents was calculated and listed in Table 1.

Antioxidant Activity Test

The DPPH scavenging activities of *D. praelonga* leaves crude extracts, fractions, pure compound and the positive control (BHT) are presented in Figure 1. The ethyl acetate and ethanol crude extracts exhibited a potential scavenging effect. However, the *n*-hexane crude extract and Fraction 1 until Fraction 8.

The decreasing scavenging activity of the samples based on the IC₅₀ was in the order; BHT > pure compound > F-9 until F-18 > ethanol crude extract > ethyl acetate crude extract > F-2 until F-8 > *n*-hexane crude extract > F-1. Content of polyphenols and flavonoids cause ability the higher antioxidant activity.

A compound that acts as an antioxidant is able to inhibit or prevent the oxidation process in a substrate that can be caused by free radicals compounds at small concentrations with a significant reduction (Isnindar et al. 2011). Based on the research conducted by Fauzi et al. (2011), compound that has a potential power of antioxidant can inhibit the growth of breast cancer with EC₅₀ values of 2.4 - 2.8%. Reported by Fitrilia et al. (2015) water extract of clove mistletoe had DPPH free scavenging activity with IC₅₀ 11.4 μg mL⁻¹. Ethanol extract *D. petandra* showed DPPH scavenging activity with value IC₅₀ 4.74 μg mL⁻¹ (Widowati, 2013).

Table 1. Extraction yield of *D. praelonga* leaves

<table>
<thead>
<tr>
<th>Sample</th>
<th>Yield (g)</th>
<th>Rendemen (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>n-Hexane extract</td>
<td>14.22</td>
<td>1.67</td>
</tr>
<tr>
<td>Ethyl acetate extract</td>
<td>16.21</td>
<td>1.90</td>
</tr>
<tr>
<td>Ethanol 96% extract</td>
<td>85.71</td>
<td>10.08</td>
</tr>
</tbody>
</table>

Figure 1. Antioxidant Activity

Figure 2. The data is expressed as percentage of antioxidant activity ± SEM, as compared to the negative control (100%). Level of significance is *P* < 0.05.
The antioxidant activity quantified by DPPH in each of the study samples, the lowest value was obtained for \textit{n}-hexane extract and followed fractions (F-1 to F-8 and F-19 to F-21) also showed no active as antioxidant (IC\textsubscript{50} > 100 μg mL\textsuperscript{-1}) (P>0,05) (Figure 2). The polarity of solvent alters its extraction effect for particular group antioxidant and the influences the scavenging capacity of extract. The \textit{ethanol} extract and \textit{ethyl acetate} extract with the highest values followed fractions (F-9 to F-18), isolated compound and positive control (BHT) respectively. Showed the significant radical scavenging activity with IC\textsubscript{50} < 50 μg mL\textsuperscript{-1} (P<0,05). DPPH colour was the fading in concentration dependent manner by \textit{D. praelonga} crude extracts, antioxidant activity increased significant (P<0,05) with increasing concentration. The antioxidant activity of \textit{D. praelonga} crude extracts varied the solvents use for extraction using maseration method.

**Isolation and Purification of Ethanol Extract**

The fractionation of \textit{ethanol} extract obtained 21 fractions. It was conducted by using column chromatography techniques in the stationary phase of silica gel, and eluent \textit{n}-hexane, ethylacetate and methanol gradient with a ratio of 10%. The next stage was an TLC analysis of each fraction produced. TLC analysis is shown in Figure 3.

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**Figure 3.** Result of chromatogram profil TLC of fractions

**Figure 4.** Result of chromatogram profil TLC of pure compound

**Figure 5.** DEPT 135° spectrum

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\textit{D. praelonga} crude extracts, antioxidant activity
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Based on TLC analysis, fractions 13 and 14 have the same stain pattern and similar ability to reduce the source of radicals, so that the fraction is put together and attained a weight of 300 mg. In the next step, the combined fraction was purified by reverse chromatography using stationary phase of ODS and methanol-water eluent isocratically. From the separation technique, we obtained a spot with a pure compound stain pattern shown in Figure 4.

Determination of the chemical structure of pure compounds obtained in the form of amorphous powder in yellow with the formula C_{15}H_{9}O_{7} with a value of m/z 301 [M-H-]. The results of infrared (IR) spectra analysis provide absorption bands at wave numbers 3400 cm\(^{-1}\) (OH); 1666 cm\(^{-1}\) (C=O); 1520 cm\(^{-1}\) (C=C); 1317 cm\(^{-1}\) (C-O (ether)) and 931 cm\(^{-1}\) (aromatic). After identifying the functional groups and molecular weights of these compound, then an NMR analysis was carried out, which involved protons, carbon, and DEPT 135\(^{O}\). From carbon analysis using the DEPT 135\(^{O}\) technique, it was shown that there are five sp\(^{2}\) metin signals, and nine quaternary sp\(^{2}\) carbon. The DEPT 135\(^{O}\) spectra image is presented in Figure 5.

### Table 2. Extraction yield of *D. praelonga* leaves

<table>
<thead>
<tr>
<th>Position</th>
<th>δ C (ppm)</th>
<th>δ H (ppm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>C</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>158.3 (s)</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>137.3 (s)</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>177.4 (s)</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>162.6 (s)</td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>6.18 (1H, d, J =2,5)</td>
<td>99.3 (d)</td>
</tr>
<tr>
<td>7</td>
<td>165.6 (s)</td>
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</tr>
<tr>
<td>8</td>
<td>6.39 (1H, d, J =2,18)</td>
<td>94.4 (d)</td>
</tr>
<tr>
<td>9</td>
<td>148.8 (s)</td>
<td></td>
</tr>
<tr>
<td>10</td>
<td>104.6 (s)</td>
<td></td>
</tr>
<tr>
<td>1’</td>
<td>121.7 (s)</td>
<td></td>
</tr>
<tr>
<td>2’</td>
<td>116.3 (d)</td>
<td></td>
</tr>
<tr>
<td>3’</td>
<td>146.3 (s)</td>
<td></td>
</tr>
<tr>
<td>4’</td>
<td>148 (s)</td>
<td></td>
</tr>
<tr>
<td>5’</td>
<td>116 (d)</td>
<td></td>
</tr>
<tr>
<td>6’</td>
<td>7.64 (1H, q, J = 8,47)</td>
<td>124.2 (d)</td>
</tr>
</tbody>
</table>

**Figure 5.** DEPT 135\(^{O}\) spectrum
The results of Nuclear Magnetic of Resonance analysis of the proton (NMR) proton and carbon (1D) provide the chemical shift presented in Table 2.

The structure of compound exhibited resonances due to aromatic systems (Figure 6). The aromatic region exhibited ABX system and to a 3’ and 4’ substitution dihydroxide of ring B and a typical meta-coupled pattern for H-6 and H-8 protons of ring A and showed the presence of 15 aromatic carbon signals.

Cytotoxic Activity

Cytotoxic test of the isolated pure compound against MCF-7 breast cancer cells gave an improvement in terms of inhibition value (%) along with the increase of sample concentration. Inhibition values (%) are presented in Figure 7.

According to the chart above, the regression equation is obtained, \( y = 0.0529x + 10.292 \) with an \( IC_{50} \) value of 750.62 \( \mu \)g/mL. Based on research conducted by Zainudin & Su’ain (2015), the ethylacetate extract of D. petandra has cytotoxic behaviors against breast cancer cells (MCF-7) with \( IC_{50} \) values of 14 \( \mu \)g/mL. Flavonoid bioactive compounds have potential as a source of antioxidants that have a correlation in the process of inhibiting the growth of cancer cells. The bioactive component of flavonoids has anti-carcinogenic properties with its ability to modulate the main target in the cell cycle path. It also has the ability to trigger the process of apoptosis, can inhibit the stages of tumor cell invasion and metastasis, and influence the signaling transduction process (Sandhar et al. 2011).

Based on the data of this research, \( IC_{50} \) towards the relatively pure compounds is not toxic. The low cytotoxic power of pure compound against breast cancer cells (MCF-7) is thought to be the nature of resistance from cells to several compounds that act as anticancer agents. The triggering factors that can cause resistance are influenced by several resistant genes such as multidrug resistance protein (MDR1), multidrug resistance associated protein (MRPs), glutathione-S-transferase (GST), dihydropyrimidine dehydrogenase (DPD) and galectin (Fulda et al. 2010).

CONCLUSION

Based on analysis using LC-MS, IR and NMR, it was concluded that the isolated compound from petai mistletoe (D. praelonga) is a flavonol, quercetine (3,5,7,3’,4’petahydroxyflavone), which has the potential as an antioxidant compound with inhibition values of 9.4 \( \mu \)g/mL, and cytotoxic values of breast cancer cells (MCF-7) of 750.62 \( \mu \)g/mL.
ACKNOWLEDGMENTS

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REFERENCES


Fauzi AN, Nur Azmi MN, Yaacob NS. 2011. Tualang honey induces apoptosis and disrupts the mitochondrial membrane potential of human breast and cervical cancer cell lines. Food and Chemical Toxicology. 49(4): 871-78


